Local immune compartments are related to the severity of dextran sodium sulphate induced colitis

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Summary

Dextran sodium sulphate (DSS) induced colitis is commonly used to simulate human ulcerative colitis (UC). However, the mucosal immune responses related to the severity of disease have not been comprehensively documented. We used different concentration of DSS, induced various severities of colitis, and simultaneously examined the frequency of immune cells, antibodies and cytokine production. We found that T regulatory cells (Tregs), B cells, and IgA secretion increased on the recovery phase of mild colitis, accompanied by CD11b+ cells, interleukin (IL)-6 and tumor necrosis factor (TNF)-α accumulated mildly. While during severe and irreversible colitis, the CD11b+ cells, IL-6, and TNF-α infiltrated severely with Tregs, B cells, and IgA increased inconspicuously. These results demonstrate that Tregs, B cells, and IgA may play a significant role in maintaining the homeostasis of gut, by suppressing CD11b+ cells and the pro-inflammatory cytokines.

Keywords: DSS induced colitis, Tregs, B cells, IgA, CD11b+ cells

1. Introduction

Ulcerative colitis (UC) is an inflammatory bowel disease characterized by intestinal inflammation and mucosal damage (1). A widely used experimental model of colitis involves oral administration of dextran sulphate sodium (DSS) dissolved in drinking water. The exact mechanism through which DSS initiates colitis is unknown. One of the possible mechanisms is that DSS activates gut-associated lymphoid tissues (GALT) immune cells (2).

GALT are important for the maintenance of intestinal homeostasis, which consist of Peyer's patches, intraepithelial lymphocytes (IEL), lamina propria (LP) and mesenteric lymph nodes (MLN). The GALT that contain lymphocytes can be divided into two main immune-function-based compartments, the bacterial and antigen elimination required effector compartment and self-tolerance regulator compartment, which are associated with colitis symptoms (3). Granulocytes and macrophages produce proinflammatory cytokines such as IL-6, TNF-α and are reported to be directly involved in the pathogenesis of UC (4,5). On the contrary, B cells are mainly located within the Peyer's patches and after cell differentiation, IgA plasmablasts migrate to the lamina propria and secrete IgA into the gut lumen (6-9). In addition, Treg cells also play an essential role in mucosal immune tolerance (10,11). The other essential protective cytokines related to B cells and Tregs are transforming growth factor (TGF)-β and IL-10 (12-16). Although the etiology and pathogenesis of colitis are still unclear, inappropriate activation of the GALT immune system has played an important role in the pathogenesis of mucosal inflammation (3). However, whether the expression of those GALT immune compartments are involved in the degree and the recovery of colitis have not yet been completely identified.

In this study, we used 1% DSS and 3.5% DSS to induce mild and severe colitis respectively. By comparing the immune compartments in the GALT during mild or severe colitis, we demonstrated that regulators such as Tregs, B cells, and IgA increased in the recovery phase of mild colitis. While during severe colitis, the amount of GALT regulators did not change or decreased which resulted in irreversible colitis.
2. Materials and Methods

2.1. Mice

Wild-type (WT) C57BL/6 mice were obtained from the Chinese Academy of Science (Shanghai, China). All mice used were 8-12 weeks of age and were housed in the animal facility of Fudan University, China. All animals received care in accordance with the Animal Care and Use Committee at Fudan University.

2.2. DSS-induced colitis

Mice were given 1% DSS (molecular weight 36-50 kDa) (MP Biomedicals, Inc., Santa Ana, CA, USA) or 3.5% DSS in their drinking water for 7 days followed by regular drinking water (17). The Disease activity index (DAI) was scored on a grade from 0 to 4 based on daily assessment of body weight, stool consistency, and detection of rectal bleeding, and then averaged for each parameter. Parameters were scored as follows: body weight loss scores (0, none; 1, 1-5%; 2, 6-10%; 3, 11-15%; 4, over 15%), stool scores (0, normal stools; 2, loose stools; 4, diarrhea) and fecal bleeding scores (0, negative hemoccult test; 1, positive hemoccult test; 2, blood visibly present in the stool; 3, blood visibly and blood clotting on the anus; 4, gross bleeding) as previously described (17).

2.3. Histology

At day 7 and day 10 after DSS administration, mice were sacrificed, the colon was removed and the segments were fixed in 10% buffered formalin. After paraffin embedding, five-micrometer thick sections were cut transversely and stained with hematoxylin and eosin (H&E). Assessment of histological colon injury was performed at 200× as previously described (18). A histology score was generated on a grade from 0 to 4 as follows: 0, normal tissue; 1, mild inflammation with scattered inflammatory cells; 2, moderate inflammation with multiple foci and/or mild epithelial ulcerations; 3, severe inflammation with massive infiltrating mononuclear cells and/or ulcerations in more than 30% of the tissue; and 4, inflammation with transmural inflammatory cell infiltration and/or more than 75% of the tissue section affected.

2.4. Gut cell isolation

MLN, Peyer's patches, intraepithelial lymphocytes (iEL) and lamina propria lymphocytes (LPL) were isolated by the method of Lefrancois and Lycke (19). Briefly, MLN were isolated and then the whole intestine was flushed with Ca²⁺Mg²⁺-free 100 mM HBSS (CMF) (Sigma-Aldrich, St. Louis, MO, USA). RPMI was injected into the intestine to highlight Peyer's patches, which were then collected with a fine forceps and scissors. The intestines were opened longitudinally and cut into small sections, shuck with CMF solution containing HEPES, NaHCO₃, DTT and EDTA (all from Sigma-Aldrich, St. Louis, MO, USA), and 2% fetal bovine serum (FBS) (Gibco®, Portland, OR, USA) at 37°C. IELs were then purified using discontinuous Percoll gradients (GE Healthcare, Buckinghamshire, UK). For the isolation of LPLs, intestinal pieces were digested with complete RPMI-1640 containing collagenase (Roche Applied Science, Upper Bavaria, Germany), dispase (Sigma-Aldrich, St. Louis, MO, USA) and Dnase I (Sigma-Aldrich, St. Louis, MO, USA) at 37°C. LPLs were then purified using Percoll gradients.

2.5. Flow cytometry and antibodies

Single cell suspensions from the MLN, Peyer's patches, epithelium (iEL) and LP were incubated with the 2.4G2 antibody (eBioscience, San Diego, CA, USA) to block FcR II/III receptors before staining with specific Ab. The anti-mouse antibodies B220-PE-Texas Red, CD3-PerCP-Cy7, CD4-APC-Cy7, Foxp3-APC and CD11b-Pacific Blue were purchased from eBioscience (San Diego, CA, USA). Surface and intracellular Foxp3 staining was performed using the manufacturer's recommended protocol. Cells were applied to a CyAn™ ADP Analyzer (Beckman, Coulter, Inc., Carlsbad, CA, USA) and data were analyzed using FlowJo software (Tree Star, Ashland, Or, USA).

2.6. ELISA

The intestines were isolated and flushed with 40 ml endotoxin free PBS. Mucus containing fluid was harvested and stored at −80°C until assayed. The concentration of IL-6, IL-10, TNF-α, TGF-β, IgM, and IgA was evaluated using the Mouse Ready-SET-Go! ELISA kits from eBioscience (San Diego, CA, USA) according to the manufacturer's instructions.

2.7. Statistical analysis

Multiple group comparisons were performed using one-way ANOVA, followed by Bonferroni Correction to compare two individual groups. The Spearman correlation was used to evaluate the relationship between immune factors and DAI. Statistical analysis was performed using STATA 8.0 or Prism 5.0. p values < 0.05 were considered statistically significant.

3. Results and Discussion

To induce UC assembled colitis, mice were treated with 1% or 3.5% DSS respectively, percentage change in body weight, DAI, colon length and histology of colonic sections of DSS treated and healthy animals were measured as follows.
In a previous study, phenotypic changes in immune compartments associated with DSS-induced colitis were determined within splenic cells and MLN. They showed that adaptive immune responses are induced during colitis (21). Thus, we asked whether the local GALT immune cells are recruited and activated during mild (1% DSS treated) or severe colitis (3.5% DSS treated). GALT cells were isolated and analyzed by flow cytometry.

On day 12, the recovery phase, there was a significant increase in the percentage of Tregs in the MLN, Peyers’ patch, iEL and LPL of 1% DSS treated mice while the frequencies didn’t alter in the GALT of 3.5% DSS treated mice. During the 1% DSS treatment, the frequencies of B cells increased in the initial compartment MLN and effected the iEL compartment of GALT while the frequencies decreased in the MLN, Peyers’ patch and LPL of 3.5% DSS treated mice (Figure 2).

On the other hand, the effector compartment of GALT-CD11b+ cells were significantly increased in the MLN, Peyers’ patch, and iEL during 3.5% DSS induced colitis while the percentage changed mildly in 1% DSS treated mice. At sites of intestinal inflammation, CD11b+ cells such as granulocytes and macrophages produced high levels of inflammatory cytokines and chemokines, contributing to the recruitment of immune cells to the inflamed tissue. These findings suggest that the severity of disease related to the mucosal immune responses.

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Figure 2. Percentage of immune cell populations in GALT on day 12. Colitis was induced as in Figure 1. Flow cytometry was used to determine the percentage of CD3+ T cells, B220+ B cells, CD11b+ cells and Tregs (CD4 Foxp3+) in Peyers’ patch, iEL, MLN and LPL of colitis mice and healthy mice 12 days after colitis induction. The data shown are the mean ± SEM from one experiment with five mice and was repeated three times with similar results. *p < 0.05; **p < 0.01; ***p < 0.001.

Figure 3. DSS-induced colitis induces significant changes in cytokines and antibodies within the gut. Colitis was induced as in Figure 1. On day 12, the intestines were flushed with 20 mL endotoxin and FCS free PBS. Mucus containing fluid was harvested and the level of IL-6, TNF-α, IL-10, TGF-β, IgM and IgA in the supernatant was determined by ELISA. Each data point represents a single mouse with the mean shown. Data shown are the mean ± SEM of six mice and was repeated three times with similar results. *p < 0.05; **p < 0.01.
pro-inflammatory cytokines, including IL-6, and TNF-α, which are directly involved in the pathogenesis of UC (4,5). On the contrary, a protective role for B cells and serum Ig was described in the TCR-α−/− spontaneous model of colitis (22) and DSS induced colitis (23). And also, Tregs are considered one of the most important immune cell regulators of intestinal homeostasis (15,24-27). Thus, the deficiency of GALT B cells and Tregs during 3.5% DSS may cause the infiltration of CD11b+ cells and lead to an irreversible colitis.

We next determined cytokine and antibody production by those immune cells during mild and severe colitis. We found that IL-10, TGF-β and IgM production in the gut mucus were increased during colitis, but there was no significant difference between mild and severe colitis, however, pro-inflammatory cytokines TNF-α and IL-6 were found to be significantly higher in the severe colitis (Figure 3). Noteworthy, IgA secreted in the gut mucus decreased in the severe colitis mice compared to the mild colitis mice (Figure 3). In order to identify the effector compartment and the regulator compartment during colitis, we performed statistical analysis between immune factors and DAI. Tregs, B cells and IgA had a negative relationship with DAI, while CD11b+ cells, IL-6 and TNF-α had a positive relationship with DAI (Figure 4).

Our results indicated that Tregs, B cells, and IgA produced had a regulatory function during colitis, which can rescue colitis by confrontation of pro-inflammation cells and cytokines (Figure 3). However, during severe colitis, Tregs, B cells and IgA were exhausted in the acute phase (data not shown). They were not increased on recovery phase, thus CD11b+ cells and its secreted IL-6 and TNF-α were accumulated on lamina propria and epithelia, resulting in severe and irreversible colitis. These results give us a new prospect that patients have different degrees of colitis (28), both immune cells such as CD11b+ cells, B cells, Tregs and the immune factors produced by them are directly related to the symptoms of different degrees of colitis. Thus, the frequencies of immune cells and the level of immune factors may provide a new diagnosis of colitis. More importantly, the fact that sufficient Tregs, B cells and IgA are able to rescue colitis from pro-inflammatory factors activation while the deficiency of regulators leads to an irreversible colitis may provide innovative immune cell-based therapies for the treatment of colitis.

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Figure 4. The expression of immune cells and cytokines is related to disease symptoms. Relative frequencies of B220+ B cells, CD11b+ cells, Tregs, IL-6, TNF-α, IgA and DAI in healthy and colitis mice. Each data point represents a single mouse with the mean shown. Data were repeated three times with similar results.